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(71) Applicant (for all designated States except US): CONOR  
MEDSYSTEMS, INC. [US/US]; 1003 Hamilton Court,  
Menlo Park, CA 94025 (US).

(72) Inventor; and

(75) Inventor/Applicant (for US only): SHANLEY, John, F.  
[US/US]; 217 Yarrowborough Lane, Redwood City, CA 94061  
(US).

(74) Agent: LYNCH, Cindy, A.; VP. Intellectual Property,  
Conor Medsystems, Inc., 1003 Hamilton Court, Menlo  
Park, CA 94025 (US).

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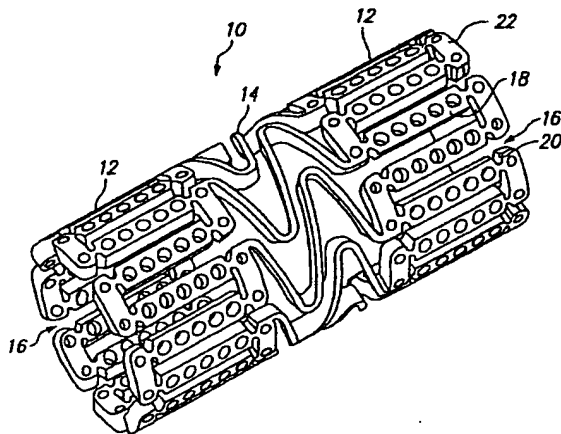
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(54) Title: EXPANDABLE MEDICAL DEVICE FOR TREATING CARDIAC ARRHYTHMIAS



(57) Abstract: An expandable medical device (10) has a plurality of elongated struts (14) joined together to form a substantially cylindrical device (12) which is expandable from a cylinder having a first diameter to a cylinder having a second diameter. At least one of the plurality of struts includes at least one opening (24, 26) extending at least partially through a thickness of said strut. A beneficial agent is loaded into the opening within the strut in layers to achieve desired temporal release kinetics of the agent. The beneficial agent can be a chemically ablative agent use to create a lesion to treat atrial fibrillation. A wide variety of delivery profiles can be achieved including zero order, pulsatile, increasing, decrease, sinusoidal, and other delivery profiles.

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## EXPANDABLE MEDICAL DEVICE FOR TREATING CARDIAC ARRHYTHMIAS

### BACKGROUND OF THE INVENTION

#### Field of the Invention

The present invention relates to tissue-supporting medical devices, and more particularly to expandable, non-removable devices that are implanted within a bodily lumen of a living animal or human to support the organ and maintain patency, and that can deliver a beneficial agent to the intervention site.

#### Summary of the Related Art

In the past, permanent or biodegradable devices have been developed for implantation within a body passageway to maintain patency of the passageway. These devices are typically introduced percutaneously, and transported transluminally until positioned at a desired location. These devices are then expanded either mechanically, such as by the expansion of a mandrel or balloon positioned inside the device, or expand themselves by releasing stored energy upon actuation within the body. Once expanded within the lumen, these devices, called stents, become encapsulated within the body tissue and remain a permanent implant.

Known stent designs include monofilament wire coil stents (U.S. Pat. No. 4,969,458); welded metal cages (U.S. Pat. Nos. 4,733,665 and 4,776,337); and, most prominently, thin-walled metal cylinders with axial slots formed around the circumference (U.S. Pat. Nos. 4,733,665; 4,739,762; and 4,776,337). Known construction materials for use in stents include polymers, organic fabrics and biocompatible metals, such as, stainless steel, gold, silver, tantalum, titanium, and shape memory alloys such as Nitinol.

U.S. Pat. Nos. 4,733,665; 4,739,762; and 4,776,337 disclose expandable and deformable interluminal vascular grafts in the form of thin-walled tubular members with axial slots allowing the members to be expanded radially outwardly into contact with a body passageway. After insertion, the tubular members are mechanically

expanded beyond their elastic limit and thus permanently fixed within the body. U.S. Pat. No. 5,545,210 discloses a thin-walled tubular stent geometrically similar to those discussed above, but constructed of a nickel-titanium shape memory alloy ("Nitinol"), which can be permanently fixed within the body without exceeding its elastic limit. All of these stents share a critical design property: in each design, the features that undergo permanent deformation during stent expansion are prismatic, i.e., the cross sections of these features remain constant or change very gradually along their entire active length. These prismatic structures are ideally suited to providing large amounts of elastic deformation before permanent deformation commences, which in turn leads to sub-optimal device performance in important properties including stent expansion force, stent recoil, strut element stability, stent securement on delivery catheters, and radiopacity.

U.S. Pat. No. 6,241,762, which is incorporated herein by reference in its entirety, discloses a non-prismatic stent design which remedies the above mentioned performance deficiencies of previous stents. In addition, preferred embodiments of this patent provide a stent with large, non-deforming strut and link elements, which can contain holes without compromising the mechanical properties of the strut or link elements, or the device as a whole. Further, these holes may serve as large, protected reservoirs for delivering various beneficial agents to the device implantation site.

Of the many problems that may be addressed through stent-based local delivery of beneficial agents, one of the most important is restenosis. Restenosis is a major complication that can arise following vascular interventions such as angioplasty and the implantation of stents. Simply defined, restenosis is a wound healing process that reduces the vessel lumen diameter by extracellular matrix deposition and vascular smooth muscle cell proliferation, and which may ultimately result in renarrowing or even reocclusion of the lumen. Despite the introduction of improved surgical techniques, devices and pharmaceutical agents, the overall restenosis rate is still reported in the range of 25% to 50% within six to twelve months after an angioplasty procedure. To treat this condition, additional revascularization procedures are frequently required, thereby increasing trauma and risk to the patient.

Some of the techniques under development to address the problem of restenosis include irradiation of the injury site and the use of conventional stents to deliver a variety of beneficial or pharmaceutical agents to the wall of the traumatized vessel. In the latter case, a conventional stent is frequently surface-coated with a beneficial agent (often a drug-impregnated polymer) and implanted at the angioplasty site. Alternatively, an external drug-impregnated polymer sheath is mounted over the stent and co-deployed in the vessel.

While acute outcomes from radiation therapies appeared promising initially, long term beneficial outcomes have been limited to reduction in restenosis occurring within a previously implanted stent, so-called 'in-stent' restenosis. Radiation therapies have not been effective for preventing restenosis in *de novo* lesions. Polymer sheaths that span stent struts have also proven problematic in human clinical trials due to the danger of blocking flow to branch arteries, incomplete apposition of stent struts to arterial walls and other problems. Unacceptably high levels of MACE (Major Adverse Cardiac Events that include death, heart attack, or the need for a repeat angioplasty or coronary artery bypass surgery) have resulted in early termination of clinical trials for sheath covered stents.

Conventional stents with surface coatings of various beneficial agents, by contrast, have shown promising early results. U.S. Pat. No. 5,716,981, for example, discloses a stent that is surface-coated with a composition comprising a polymer carrier and paclitaxel (a well-known compound that is commonly used in the treatment of cancerous tumors). The patent offers detailed descriptions of methods for coating stent surfaces, such as spraying and dipping, as well as the desired character of the coating itself: it should "coat the stent smoothly and evenly" and "provide a uniform, predictable, prolonged release of the anti-angiogenic factor." Surface coatings, however, can provide little actual control over the release kinetics of beneficial agents. These coatings are necessarily very thin, typically 5 to 8 microns deep. The surface area of the stent, by comparison is very large, so that the entire volume of the beneficial agent has a very short diffusion path to discharge into the surrounding tissue.

Increasing the thickness of the surface coating has the beneficial effects of improving drug release kinetics including the ability to control drug release and to

allow increased drug loading. However, the increased coating thickness results in increased overall thickness of the stent wall. This is undesirable for a number of reasons, including increased trauma to the vessel wall during implantation, reduced flow cross-section of the lumen after implantation, and increased vulnerability of the coating to mechanical failure or damage during expansion and implantation. Coating thickness is one of several factors that affect the release kinetics of the beneficial agent, and limitations on thickness thereby limit the range of release rates, durations, and the like that can be achieved.

In addition to sub-optimal release profiles, there are further problems with surface coated stents. The fixed matrix polymer carriers frequently used in the device coatings typically retain approximately 30% of the beneficial agent in the coating indefinitely. Since these beneficial agents are frequently highly cytotoxic, sub-acute and chronic problems such as chronic inflammation, late thrombosis, and late or incomplete healing of the vessel wall may occur. Additionally, the carrier polymers themselves are often highly inflammatory to the tissue of the vessel wall. On the other hand, use of bio-degradable polymer carriers on stent surfaces can result in the creation of "virtual spaces" or voids between the stent and tissue of the vessel wall after the polymer carrier has degraded, which permits differential motion between the stent and adjacent tissue. Resulting problems include micro-abrasion and inflammation, stent drift, and failure to re-endothelialize the vessel wall.

Another significant problem is that expansion of the stent may stress the overlying polymeric coating causing the coating to plastically deform or even to rupture, which may therefore effect drug release kinetics or have other untoward effects. Further, expansion of such a coated stent in an atherosclerotic blood vessel will place circumferential shear forces on the polymeric coating, which may cause the coating to separate from the underlying stent surface. Such separation may again have untoward effects including embolization of coating fragments causing vascular obstruction.

### SUMMARY OF THE INVENTION

In view of the drawbacks of the prior art, it would be advantageous to provide a stent capable of delivering a beneficial agent including an ablative agent to a site in a vessel.

In accordance with one aspect of the invention, a medical device for treating cardiac arrhythmias includes a bioresorbable implantable device, a plurality of openings in the implantable device, and a chemically ablative agent provided in the openings, wherein the openings are configured to deliver the chemically ablative agent to tissue surrounding the expandable cylindrical device without permanently trapping any agent in the openings.

In accordance with a further aspect of the invention, a method of treating atrial fibrillation includes providing a stent having a chemically ablative agent in a biodegradable matrix, determining an ablation location within a blood vessel of a patient where ablation is desired, implanting the stent at the ablation location and delivering the chemically ablative agent to the wall of the blood vessel without substantially delivering the chemically ablative agent into the vessel lumen, and creating a lesion in the blood vessel wall which electrically isolates portions of tissue.

### BRIEF DESCRIPTION OF THE DRAWINGS

The invention will now be described in greater detail with reference to the preferred embodiments illustrated in the accompanying drawings, in which like elements bear like reference numerals, and wherein:

FIG. 1 is a perspective view of a tissue supporting device in accordance with a first preferred embodiment of the present invention;

FIG. 2 is an enlarged side view of a portion of the device of FIG. 1;

FIG. 3 is an enlarged side view of a tissue supporting device in accordance with a further preferred embodiment of the present invention;

FIG. 4 is an enlarged side view of a portion of the stent shown in FIG. 3;

FIG. 5 is an enlarged cross section of an opening;

FIG. 6 is an enlarged cross section of an opening illustrating beneficial agent loaded into the opening;

FIG. 7 is an enlarged cross section of an opening illustrating a beneficial agent loaded into the opening and a thin coating of a beneficial agent;

FIG. 8 is an enlarged cross section of an opening illustrating a beneficial agent loaded into the opening and thin coatings of different beneficial agents on different surfaces of the device;

FIG. 9 is an enlarged cross section of an opening illustrating a beneficial agent provided in a plurality of layers;

FIG. 10 is an enlarged cross section of an opening illustrating a beneficial agent and a barrier layer loaded into the opening in layers;

FIG. 11A is an enlarged cross section of an opening illustrating a beneficial agent, a biodegradable carrier, and a barrier layer loaded into the opening in layers;

FIG. 11B is a graph of the release kinetics of the device of FIG. 11A;

FIG. 12 is an enlarged cross section of an opening illustrating different beneficial agents, carrier, and barrier layers loaded into the opening;

FIG. 13 is an enlarged cross section of an opening illustrating a beneficial agent loaded into the opening in layers of different concentrations;

FIG. 14 is an enlarged cross section of an opening illustrating a beneficial agent loaded into the opening in layers of microspheres of different sizes;

FIG. 15A is an enlarged cross section of a tapered opening illustrating a beneficial agent loaded into the opening;

FIG. 15B is an enlarged cross section of the tapered opening of FIG. 15A with the beneficial agent partially degraded;

FIG. 15C is a graph of the release kinetics of the device of FIGS. 15A and 15B;

FIG. 16A is an enlarged cross section of an opening illustrating a beneficial agent loaded into the opening in a shape configured to achieve a desired agent delivery profile;

FIG. 16B is an enlarged cross section of the opening of FIG. 16A with the beneficial agent partially degraded;

FIG. 16C is a graph of the release kinetics of the device of FIGS. 16A and 16B;

FIG. 17A is an enlarged cross section of an opening illustrating the beneficial agent loaded into the opening and a spherical shape;

FIG. 17B is a graph of the release kinetics of the device of FIG. 17A;

FIG. 18A is an enlarged cross section of an opening illustrating a plurality of beneficial agent layers and a barrier layer with an opening for achieving a desired agent delivery profile;

FIG. 18B is an enlarged cross section of the opening of FIG. 18A with the agent layers beginning to degraded;

FIG. 18C is an enlarged cross section of the opening of FIG. 18A with the agent layers further degraded; and

FIG. 19 is an enlarged cross section of an opening illustrating a plurality of cylindrical beneficial agent layers.

#### **DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS**

Referring to FIGS. 1 and 2, a tissue supporting device in accordance with one preferred embodiment of the present invention is shown generally by reference numeral 10. The tissue supporting device 10 includes a plurality of cylindrical tubes 12 connected by S-shaped bridging elements 14. The bridging elements 14 allow the tissue supporting device to bend axially when passing through the tortuous path of the vasculature to the deployment site and allow the device to bend when necessary to match the curvature of a vessel wall to be supported. Each of the cylindrical tubes 12 has a plurality of axial slots 16 extending from an end surface of the cylindrical tube toward an opposite end surface.

Formed between the slots 16 is a network of axial struts 18 and links 22. The struts 18 and links 22 are provided with openings for receiving and delivering a beneficial agent. As will be described below with respect to FIGS. 9-17, the beneficial agent is loaded into the openings in layers or other configurations which provide control over the temporal release kinetics of the agent.

Each individual strut 18 is preferably linked to the rest of the structure through a pair of reduced sections 20, one at each end, which act as stress/strain concentration features. The reduced sections 20 of the struts function as hinges in the cylindrical structure. Since the stress/strain concentration features are designed



to operate into the plastic deformation range of generally ductile materials, they are referred to as ductile hinges 20. The ductile hinges 20 are described in further detail in U.S. Pat. No. 6,241,762, which has been incorporated herein by reference.

With reference to the drawings and the discussion, the *width* of any feature is defined as its dimension in the circumferential direction of the cylinder. The *length* of any feature is defined as its dimension in the axial direction of the cylinder. The *thickness* of any feature is defined as the wall thickness of the cylinder.

The presence of the ductile hinges 20 allows all of the remaining features in the tissue supporting device to be increased in width or the circumferentially oriented component of their respective rectangular moments of inertia - thus greatly increasing the strength and rigidity of these features. The net result is that elastic, and then plastic deformation commence and propagate in the ductile hinges 20 before other structural elements of the device undergo any significant elastic deformation. The force required to expand the tissue supporting device 10 becomes a function of the geometry of the ductile hinges 20, rather than the device structure as a whole, and arbitrarily small expansion forces can be specified by changing hinge geometry for virtually any material wall thickness. The ability to increase the width and thickness of the struts 18 and links 22 provides additional area and depth for the beneficial agent receiving openings.

In the preferred embodiment of FIGS. 1 and 2, it is desirable to increase the width of the individual struts 18 between the ductile hinges 20 to the maximum width that is geometrically possible for a given diameter and a given number of struts arrayed around that diameter. The only geometric limitation on strut width is the minimum practical width of the slots 16 which is about 0.002 inches (0.0508 mm) for laser machining. Lateral stiffness of the struts 18 increases as the cube of strut width, so that relatively small increases in strut width significantly increase strut stiffness. The net result of inserting ductile hinges 20 and increasing strut width is that the struts 18 no longer act as flexible leaf springs, but act as essentially rigid beams between the ductile hinges. All radial expansion or compression of the cylindrical tissue supporting device 10 is accommodated by mechanical strain in the hinge features 20, and yield in the hinge commences at very small overall radial

expansion or compression.

The ductile hinge 20 illustrated in FIGS. 1 and 2 is exemplary of a preferred structure that will function as a stress/strain concentrator. Many other stress/strain concentrator configurations may also be used as the ductile hinges in the present invention, as shown and described by way of example in U.S. Pat. No. 6,241,762. The geometric details of the stress/strain concentration features or ductile hinges 20 can be varied greatly to tailor the exact mechanical expansion properties to those required in a specific application.

Although a tissue supporting device configuration has been illustrated in FIG. 1 which includes ductile hinges, it should be understood that the beneficial agent may be contained in openings in stents having a variety of designs including the designs illustrated in U.S. Provisional Patent Application Serial No. 60/314,360, filed on August 20, 2001 and U.S. Patent Application Serial No. 09/948,987, filed on September 7, 2001, which are incorporated herein by reference. The present invention incorporating beneficial agent openings may also be used with other known stent designs.

As shown in FIGS. 1-4, at least one and more preferably a series of openings 24 are formed by laser drilling or any other means known to one skilled in the art at intervals along the neutral axis of the struts 18. Similarly, at least one and preferably a series of openings 26 are formed at selected locations in the links 22. Although the use of openings 24 and 26 in both the struts 18 and links 22 is preferred, it should be clear to one skilled in the art that openings could be formed in only one of the struts and links. Openings may also be formed in the bridging elements 14. In the embodiment of FIGS. 1 and 2, the openings 24, 26 are circular in nature and form cylindrical holes extending through the width of the tissue supporting device 10. It should be apparent to one skilled in the art, however, that openings of any geometrical shape or configuration could of course be used without departing from the scope of the present invention. In addition, openings having a depth less than the thickness of the device may also be used.

The behavior of the struts 18 in bending is analogous to the behavior of an I-beam or truss. The outer edge elements 32 of the struts 18, shown in FIG. 2, correspond to the I-beam flange and carry the tensile and compressive stresses,

whereas the inner elements 34 of the struts 18 correspond to the web of an I-beam which carries the shear and helps to prevent buckling and wrinkling of the faces. Since most of the bending load is carried by the outer edge elements 32 of the struts 18, a concentration of as much material as possible away from the neutral axis results in the most efficient sections for resisting strut flexure. As a result, material can be judiciously removed along the axis of the strut so as to form openings 24, 26 without adversely impacting the strength and rigidity of the strut. Since the struts 18 and links 22 thus formed remain essentially rigid during stent expansion, the openings 24, 26 are also non-deforming.

The openings 24, 26 in the struts 18 may promote the healing of the intervention site by promoting regrowth of the endothelial cells. By providing the openings 24, 26 in the struts, 18, the cross section of the strut is effectively reduced without decreasing the strength and integrity of the strut, as described above. As a result, the overall distance across which endothelial cell regrowth must occur is also reduced to approximately 0.0025 - 0.0035 inches, which is approximately one-half of the thickness of a conventional stent. It is further believed that during insertion of the expandable medical device, cells from the endothelial layer may be scraped from the inner wall of the vessel by the openings 24, 26 and remain therein after implantation. The presence of such endothelial cells would thus provide a basis for the healing of the vessel wall.

The openings 24, 26 are loaded with an agent, most preferably a beneficial agent, for delivery to the vessel wall which the tissue supporting device 10 is supporting.

The terms "agent" and "beneficial agent" as used herein are intended to have their broadest possible interpretation and are used to include any therapeutic agent or drug, as well as inactive agents such as barrier layers or carrier layers. The terms "drug" and "therapeutic agent" are used interchangeably to refer to any therapeutically active substance that is delivered to a bodily conduit of a living being to produce a desired, usually beneficial, effect. The present invention is particularly well suited for the delivery of antiproliferatives (anti-restenosis agents) such as paclitaxel and rapamycin for example, and antithrombins such as heparin, for example.

The beneficial agents used in the present invention include classical small molecular weight therapeutic agents commonly referred to as drugs including all classes of action as exemplified by, but not limited to: antiproliferatives, antithrombins, antiplatelet, antilipid, anti-inflammatory, and anti-angiogenic, vitamins, ACE inhibitors, vasoactive substances, antimetabolites, metallo-proteinase inhibitors, NO donors, estradiols, anti-sclerosing agents, alone or in combination. Beneficial agent also includes larger molecular weight substances with drug like effects on target tissue sometimes called biologic agents including but not limited to: peptides, lipids, protein drugs, enzymes, oligonucleotides, ribozymes, genetic material, prions, virus, bacteria, and eucaryotic cells such as endothelial cells, monocyte/macrophages or vascular smooth muscle cells to name but a few examples. Other beneficial agents may include but not be limited to physical agents such as microspheres, microbubbles, liposomes, radioactive isotopes, or agents activated by some other form of energy such as light or ultrasonic energy, or by other circulating molecules that can be systemically administered.

The embodiment of the invention shown in FIGS. 1 and 2 can be further refined by using Finite Element Analysis and other techniques to optimize the deployment of the beneficial agent within the openings of the struts and links. Basically, the shape and location of the openings 24, 26 can be modified to maximize the volume of the voids while preserving the relatively high strength and rigidity of the struts 18 with respect to the ductile hinges 20.

FIG. 3 illustrates a further preferred embodiment of the present invention, wherein like reference numerals have been used to indicate like components. The tissue supporting device 100 includes a plurality of cylindrical tubes 12 connected by S-shaped bridging elements 14. Each of the cylindrical tubes 12 has a plurality of axial slots 16 extending from an end surface of the cylindrical tube toward an opposite end surface. Formed between the slots 16 is a network of axial struts 18 and links 22. Each individual strut 18 is linked to the rest of the structure through a pair of ductile hinges 20, one at each end, which act as stress/strain concentration features. Each of the ductile hinges 20 is formed between an arc surface 28 and a concave notch surface 29.

At intervals along the neutral axis of the struts 18, at least one and more preferably a series of openings 24' are formed by laser drilling or any other means known to one skilled in the art. Similarly, at least one and preferably a series of openings 26' are formed at selected locations in the links 22. Although the use of openings 24', 26' in both the struts 18 and links 22 is preferred, it should be clear to one skilled in the art that openings could be formed in only one of the struts and links. In the illustrated embodiment, the openings 24' in the struts 18 are generally rectangular whereas the openings 26' in the links 22 are polygonal. It should be apparent to one skilled in the art, however, that openings of any geometrical shape or configuration could of course be used, and that the shape of openings 24, 24' may be the same or different from the shape of openings 26, 26', without departing from the scope of the present invention. As described in detail above, the openings 24', 26' may be loaded with an agent, most preferably a beneficial agent, for delivery to the vessel in which the tissue support device 100 is deployed. Although the openings 24', 26' are preferably through openings, they may also be recesses extending only partially through the thickness of the struts and links.

The relatively large, protected openings 24, 24', 26, 26', as described above, make the expandable medical device of the present invention particularly suitable for delivering agents having more esoteric larger molecules or genetic or cellular agents, such as, for example, protein drugs, enzymes, antibodies, antisense oligonucleotides, ribozymes, gene/vector constructs, and cells (including but not limited to cultures of a patient's own endothelial cells). Many of these types of agents are biodegradable or fragile, have a very short or no shelf life, must be prepared at the time of use, or cannot be pre-loaded into delivery devices such as stents during the manufacture thereof for some other reason. The large through-openings in the expandable device of the present invention form protected areas or receptors to facilitate the loading of such an agent either at the time of use or prior to use, and to protect the agent from abrasion and extrusion during delivery and implantation.

The volume of beneficial agent that can be delivered using through openings is about 3 to 10 times greater than the volume of a 5 micron coating covering a stent with the same stent/vessel wall coverage ratio. This much larger beneficial agent capacity provides several advantages. The larger capacity can be used to deliver

multi-drug combinations, each with independent release profiles, for improved efficacy. Also, larger capacity can be used to provide larger quantities of less aggressive drugs and to achieve clinical efficacy without the undesirable side-effects of more potent drugs, such as retarded healing of the endothelial layer.

Through openings also decrease the surface area of the beneficial agent bearing compounds to which the vessel wall surface is exposed. For typical devices with beneficial agent openings, this exposure decreases by a factors ranging from about 6:1 to 8:1, by comparison with surface coated stents. This dramatically reduces the exposure of vessel wall tissue to polymer carriers and other agents that can cause inflammation, while simultaneously increasing the quantity of beneficial agent delivered, and improving control of release kinetics.

FIG. 4 shows an enlarged view of one of the struts 18 of device 100 disposed between a pair of ductile hinges 20 having a plurality of openings 24'. FIG. 5 illustrates a cross section of one of the openings 24' shown in FIG. 4. FIG. 6 illustrates the same cross section when a beneficial agent 36 has been loaded into the opening 24' of the strut 18. Optionally, after loading the opening 24' and/or the opening 26' with a beneficial agent 36, the entire exterior surface of the stent can be coated with a thin layer of a beneficial agent 38, which may be the same as or different from the beneficial agent 36, as schematically shown in FIG. 7. Still further, another variation of the present invention would coat the outwardly facing surfaces of the stent with a first beneficial agent 38 while coating the inwardly facing surfaces of the stent with a different beneficial agent 39, as illustrated in FIG. 8. The inwardly facing surface of the stent would be defined as at least the surface of the stent which, after expansion, forms the inner passage of the vessel. The outwardly facing surface of the stent would be defined as at least the surface of the stent which, after expansion, is in contact with and directly supports the inner wall of the vessel. The beneficial agent 39 coated on the inner surfaces may be a barrier layer which prevents the beneficial agent 36 from passing into the lumen of the blood vessel and being washed away in the blood stream.

FIG. 9 shows a cross section of an opening 24 in which one or more beneficial agents have been loaded into the opening 24 in discrete layers 50. One method of creating such layers is to deliver a solution comprising beneficial agent.

polymer carrier, and a solvent into the opening and evaporating the solvent to create a thin solid layer of beneficial agent in the carrier. Other methods of delivering the beneficial agent can also be used to create layers. According to another method for creating layers, a beneficial agent may be loaded into the openings alone if the agent is structurally viable without the need for a carrier. The process can then be repeated until each opening is partially or entirely filled.

In a typical embodiment, the total depth of the opening 24 is about 125 to about 140 microns, and the typical layer thickness would be about 2 to about 50 microns, preferably about 12 microns. Each typical layer is thus individually about twice as thick as the typical coating applied to surface-coated stents. There would be at least two and preferably about ten to twelve such layers in a typical opening, with a total beneficial agent thickness about 25 to 28 times greater than a typical surface coating. According to one preferred embodiment of the present invention, the openings have an area of at least  $5 \times 10^{-6}$  square inches, and preferably at least  $7 \times 10^{-6}$  square inches.

Since each layer is created independently, individual chemical compositions and pharmacokinetic properties can be imparted to each layer. Numerous useful arrangements of such layers can be formed, some of which will be described below. Each of the layers may include one or more agents in the same or different proportions from layer to layer. The layers may be solid, porous, or filled with other drugs or excipients.

FIG. 9 shows the simplest arrangement of layers including identical layers 50 that together form a uniform, homogeneous distribution of beneficial agent. If the carrier polymer were comprised of a biodegradable material, then erosion of the beneficial agent containing carrier would occur on both faces of the opening at the same time, and beneficial agent would be released at an approximately linear rate over time corresponding to the erosion rate of the carrier. This linear or constant release rate is referred to as a zero order delivery profile. Use of biodegradable carriers in combination with through openings is especially useful, to guarantee 100% discharge of the beneficial agent within a desired time without creating virtual spaces or voids between the radially outermost surface of the stent and tissue of the vessel wall. When the biodegradable material in the through openings is removed,

the openings may provide a communication between the strut-covered vessel wall and the blood stream. Such communication may accelerate vessel healing and allow the ingrowth of cells and extracellular components that more thoroughly lock the stent in contact with the vessel wall. Alternatively, some through-openings may be loaded with beneficial agent while others are left unloaded. The unloaded holes could provide an immediate nidus for the ingrowth of cells and extracellular components to lock the stent into place, while loaded openings dispense the beneficial agent.

The advantage of complete erosion using the through openings over surface coated stents opens up new possibilities for stent-based therapies. In the treatment of cardiac arrhythmias, such as atrial fibrillation both sustained and paroxysmal, sustained ventricular tachycardia, super ventricular tachycardia including reentrant and ectopic, and sinus tachycardia, a number of techniques under development attempt to ablate tissue in the pulmonary veins or some other critical location using various energy sources, e.g. microwaves, generally referred to as radio-frequency ablation, to create a barrier to the propagation of undesired electrical signals in the form of scar tissue. Focal atrial fibrillation ablation procedures use a radio-frequency catheter to electrically isolate pulmonary vein foci from the rest of the left atrium by creation of circular scars in the pulmonary veins. These techniques have proven difficult to control accurately.

A stent based therapy using through openings, biodegradable carriers, and associated techniques described herein can be used to deliver a *chemically* ablative agent in a specific, precise pattern to a specific area for treatment of atrial fibrillation, while guaranteeing that none of the inherently cytotoxic ablating agent could be permanently trapped in contact with the tissue of the vessel wall. The stent design for treatment of cardiac arrhythmias can have a relatively short length, such as 10mm or less, which can be sufficient to create the desired circular lesion. Further, the chemically ablative agent can be provided only on a short segment of the stent whether the stent is long or short. A longer stent can be used for improved stability. When a segment shorter than the full length of the stent is provided with the agent, marker bands can be provided on or in the catheter to clearly indicate the boundaries of the agent containing portion of the stent during a procedure. The



marker bands can identify to the practitioner the precise location of the agent to be delivered.

Chemically ablative agents are those which can create a refractory scar or barrier which prevents electrical impulse propagation through the pulmonary veins. Chemically ablative agents include without limitation those agents used as chemotherapeutic agents, sclerosing agents, and agents which ablate or otherwise injure tissue to create a controlled scar or tissue injury.

Examples of chemically ablative agents include without limitation, phenolic compounds, such as, phenol, cresol (o-, m- or p-), resorcinol, alkyl resorcinols (e.g. hexyl resorcinol), pyrogallol; organic acid compounds, such as, acetic acid, propionic acid, butyric acid, capric acid, caprylic acid, lactic acid, glycolic (hydroxy acetic) acid, haloalkyl acids (e.g. chloroacetic acid, dichloroacetic acid, trichloroacetic acid), citric acid, tartaric acid, ascorbic acid, glutamic acid, glutaric acid, benzoic acid, halo benzoic acids (e.g. chlorobenzoic acid), nitro benzoic acids (e.g. picric acid); hydroxy benzoic acids and derivatives, such as, salicylic acid, gallic acid, hydroxybenzoic acid, and acid ester derivatives; aldehyde compounds, such as formaldehyde, glutaraldehyde; hydroxy urea; amino saccharides, such as, streptomycin; guanidino saccharides, such as, gentamicin; ribosome inactivating proteins, such as saporin; dehydrating agents; and sclerosing agents, such as a mixture of formalin / iodine / chlorhexidine / silver nitrate, e.g. in 10 / 2 / 2 / 2 ratio. Other agents which can be used as chemically ablative agents are those that form a scar by enzymatic degradation (pepsin, trypsin, and other proteases) or oxidative degradation (peroxides).

The stent containing a chemically ablative agent can be used to create lesions in the shape of circumferential rings at a location specifically determined by a mapping procedure. The chemically ablative agents can remove or otherwise injure tissue to create a lesion.

The stent itself can also form the scar tissue or refractory tissue which prevents electrical signal propagation. For example, some stents materials can cause restenosis, a type of scarring, which can block the transmission impulses from within the pulmonary vein. Other bioresorbable stents can create a scar like deposit of calcium and/or other materials as they degrade. This deposit which remains after the

stent is resorbed can prevent electrical signal propagation.

In treatment of areas such as the pulmonary veins an entirely bioresorbable stent can be used with a first bioresorbable material forming the structural stent and another bioresorbable material forming the matrix containing the chemically ablative agent within the openings. The use of an entirely bioresorbable stent addresses any possible concerns relating to the long term presence of a metal or other material in the pulmonary veins or other vessel. The bioresorbable structure of the anti-arrhythmia stent can be formed of a bioresorbable metal alloy or a bioresorbable polymer. Bioresorbable metal alloys useful for stents include zinc-titanium alloys, and magnesium alloys, such as lithium-magnesium, sodium-magnesium, and magnesium alloys containing rare earth metals. Some examples of bioresorbable metal alloys are described in U.S. Patent No. 6,287,332, which is incorporated herein by reference in its entirety. Bioresorbable metal alloy stents can be formed in the configurations illustrated herein by laser cutting. When cutting stents from these alloys, an inert atmosphere may be desired to minimize oxidation of the alloy during cutting in which case, a helium gas stream, or other inert atmosphere can be applied during cutting. Magnesium alloys are used in the aeronautic industry and the processing systems used for the aeronautic industry can also be used for forming the stents.

Bioresorbable metal alloys provide the necessary structural strength needed for the stent, however, it is difficult to incorporate a drug within the bioresorbable metal alloy and is difficult to release the drug if it could be incorporated. More importantly, the use of drug coatings on the bioresorbable metal alloy surface can interfere with the biodegradation of the stent. Therefore, the combination of openings in the bioresorbable stent filled with a bioresorbable matrix containing chemically ablative agent provides a stent which can create a electrically refractory scar and then can disappear from the vessel.

The bioresorbable anti-arrhythmia stent provides a solution to the problem of coating a bioresorbable stent by selecting a first bioresorbable polymer for the struts of the stent and providing openings in the stent containing a beneficial agent matrix. The polymer or other matrix material in the openings requires none of the structural properties of the stent, and also requires very little flexibility or adhesion which is

required by a coating. Thus, the matrix material selection may be made based on the ability of the material to release the drug with a desired release profile. Directional delivery of one or more drugs can also be achieved with reservoirs which cannot be easily achieved with coatings, impregnation, or other methods.

Examples of bioresorbable polymers which can be used for the structural struts of the stent 10 include, without limitation, polylactic acid (PLA), polyglycolic acid (PGA), copolymers of PLA and PGA, poly-L-lactide (PLLA), poly-D,L-lactide (PDLA), poly-ε-caprolactone (PCL), and combinations thereof. U.S. Patent No. 4,889,119, which is incorporated herein by reference in its entirety, describes some of the bioresorbable polymers which are useful in the present invention.

Examples of bioresorbable polymers which can be used for the polymer/drug matrix within the reservoirs include, without limitation, polylactic acid (PLA); polyglycolic acid (PGA); copolymers of PLA and PGA; polylactic-co-glycolic acid (PLGA); poly-L-lactide (PLLA); poly-D,L-lactide (PDLA); poly-ε caprolactone (PCL); polyethylene glycol and polyethylene oxide, poly (block-ethylene oxide-block-propylene oxide-block-ethylene oxide); polyvinyl pyrrolidone; polyorthoesters; polysaccharides and polysaccharide derivatives such as polyhyaluronic acid, poly (glucose), polyalginic acid, chitin, chitosan, chitosan derivatives, cellulose, methyl cellulose, hydroxyethylcellulose, hydroxypropylcellulose, carboxymethylcellulose, cyclodextrins and substituted cyclodextrins, such as beta-cyclodextrin sulfobutyl ethers; polypeptides and proteins, such as polylysine, polyglutamic acid, albumin; and combinations thereof. Preferably, the polymer in the reservoir degrades at a rate which results in degradation of the matrix substantially at the same time or before the degradation of the stent itself.

Bioresorbable polymer stents can be formed by known methods including molding, extrusion, other thermoforming processes, laser cutting, semiconductor fabrication methods including microdischarge machining or a combination of these processes. These methods are described further in U.S. Patent Application Serial No. 10/822,063, filed on April 8, 2004, which is incorporated herein by reference in its entirety. The stent illustrated herein is only one example of the type of stent structure which may be made. Many other stent configurations can also be used

including woven stents, coil stents, serpentine patterns, diamond patterns, chevron or other patterns, or racheting or locking stents.

The anti-arrhythmia stent provides short term delivery of a potent chemically ablative agent. The agent is delivered over a short time, such as less than 30 days, while the stent can be bioresorbable in a short time such as six months, three months or less.

If, on the other hand, the goal of a particular therapy is to provide a long term effect, beneficial agents located in openings provide an equally dramatic advantage over surface coated devices. In this case, a composition comprising a beneficial agent and a non-biodegradable carrier would be loaded into the through openings, preferably in combination with a diffusion barrier layer as described below. To continue the cardiac arrhythmias example, it might be desirable to introduce a long-term anti-arrhythmic drug near the ostia of the pulmonary veins or some other critical location.

In this alternative anti-arrhythmic stent the anti-arrhythmic drugs are delivered over a long period of time directly to the pulmonary veins or other location of propagation of irregular electrical impulses. In general "anti-arrhythmic" (anti irregular heart rhythm) drugs are grouped in "classes" according to how they work. Class I are Sodium Channel Blockers which decrease the speed of electrical conduction in the heart muscle. Class II are Beta-Adrenergic or Beta-Blockers which slow down conduction through the heart and make the AV node less sensitive to irregular impulses. Class III are Potassium Channel Blockers which slow nerve impulses in the heart. Class IV are Calcium Channel Blockers which prevent or slow the flow of calcium ions into smooth muscle cells such as the heart. This impedes muscle cell contraction, thereby allowing blood vessels to expand and carry more blood and oxygen to tissues. Examples of Anti-arrhythmic drugs include Procainamide, Quinidine, Disopyramide, Flecainide, Sotalol, Dofetilide, Amiodarone, and Ibutilid.

The ability of through openings to deliver a beneficial agent, such as an anti-arrhythmic drug in a controlled manner over a long period of time provides a significant advantage over surface coatings. The transient diffusion behavior of a beneficial agent through a non-biodegradable carrier matrix can be generally

described by Fick's second law:

$$\frac{\partial C}{\partial t} = \frac{\partial}{\partial x} \left[ D \frac{\partial C}{\partial x} \right]$$

Where C is the concentration of beneficial agent at cross section x, x is either the thickness of a surface coating or depth of a through opening, D is the diffusion coefficient and t is time. The solution of this partial differential equation for a through opening with a barrier layer will have the form of a *normalized probability integral* or *Gaussian Error Function*, the argument of which will contain the term:

$$\frac{x}{2\sqrt{Dt}}$$

To compare the time intervals over which a given level of therapy can be sustained for surface coatings vs. through openings, we can use Fick's Second Law to compare the times required to achieve equal concentrations at the most inward surfaces of the coating and opening respectively, i.e. the values of x and t for which the arguments of the Error Function are equal:

$$\frac{x_1}{2\sqrt{Dt_1}} = \frac{x_2}{2\sqrt{Dt_2}} \Rightarrow \frac{x_1^2}{x_2^2} = \frac{t_1}{t_2}$$

The ratio of diffusion times to achieve comparable concentrations thus varies as the square of the ratio of depths. A typical opening depth is about 140 microns while a typical coating thickness is about 5 micron; the square of this ratio is 784, meaning that the effective duration of therapy for through openings is potentially almost three orders of magnitude greater for through openings than for surface coatings of the same composition. The inherent non-linearity of such release profiles can in part be

compensated for in the case of through openings, but not in thin surface coatings, by varying the beneficial agent concentration of layers in a through opening as described below. It will be recalled that, in addition to this great advantage in beneficial agent delivery *duration*, through openings are capable of delivering a 3 to 10 times greater *quantity* of beneficial agent, providing a decisive overall advantage in sustained therapies. The diffusion example above illustrates the general relationship between depth and diffusion time that is characteristic of a wider class of solid state transport mechanisms.

Beneficial agent that is released to the radially innermost or inwardly facing surface known as the lumen facing surface of an expanded device may be rapidly carried away from the targeted area, for example by the bloodstream, and thus lost. Up to half of the total agent loaded in such situations may have no therapeutic effect due to being carried away by the bloodstream. This is probably the case for all surface coated stents as well as the through opening device of FIG. 9.

FIG. 10 shows a device in which the first layer 52 is loaded into a through opening 24 such that the inner surface of the layer is substantially co-planar with the inwardly facing surface 54 of the cylindrical device. The first layer 52 is comprised of a material called a barrier material which blocks or retards biodegradation of subsequent layers in the inwardly facing direction toward the vessel lumen, and/or blocks or retards diffusion of the beneficial agent in that direction. Biodegradation of other layers or beneficial agent diffusion can then proceed only in the direction of the outwardly facing surface 56 of the device, which is in direct contact with the targeted tissue of the vessel wall. The barrier layer 52 may also function to prevent hydration of inner layers of beneficial agent and thus prevent swelling of the inner layers when such layers are formed of hygroscopic materials. The barrier layer 52 may further be comprised of a biodegradable material that degrades at a much slower rate than the biodegradable material in the other layers, so that the opening will eventually be entirely cleared. Providing a barrier layer 52 in the most inwardly facing surface of a through-opening thus guarantees that the entire load of beneficial agent is delivered to the target area in the vessel wall. It should be noted that providing a barrier layer on the inwardly facing surface of a surface-coated stent without openings *does not* have the same effect: since the beneficial agent in such a

coating cannot migrate through the metal stent to the target area on the outer surface, it simply remains trapped on the inner diameter of the device, again having no therapeutic effect.

Barrier layers can be used to control beneficial agent release kinetics in more sophisticated ways. A barrier layer 52 with a pre-determined degradation time could be used to deliberately terminate the beneficial agent therapy at a pre-determined time, by exposing the underlying layers to more rapid bio-degradation from both sides. Barrier layers can also be formulated to be activated by a separate, systemically applied agent. Such systemically applied agent could change the porosity of the barrier layer and/or change the rate of bio-degradation of the barrier layer or the bulk beneficial agent carrier. In each case, release of the beneficial agent could be activated by the physician at will by delivery of the systemically applied agent. A further embodiment of physician activated therapy would utilize a beneficial agent encapsulated in micro-bubbles and loaded into device openings. Application of ultrasonic energy from an exterior of the body could be used to collapse the bubbles at a desired time, releasing the beneficial agent to diffuse to the outwardly facing surface of the reservoirs. These activation techniques can be used in conjunction with the release kinetics control techniques described herein to achieve a desired drug release profile that can be activated and/or terminated at selectable points in time.

FIG. 11A shows an arrangement of layers provided in a through opening in which layers 50 of a beneficial agent in a biodegradable carrier material, are alternated with layers 58 of the biodegradable carrier material alone, with no active agent loaded, and a barrier layer 52 is provided at the inwardly facing surface. As shown in the release kinetics plot of FIG. 11B, such an arrangement releases beneficial agent in three programmable bursts or waves achieving a stepped or pulsatile delivery profile. The use of carrier material layers without active agent creates the potential for synchronization of drug release with cellular biochemical processes for enhanced efficacy.

Alternatively, different layers could be comprised of different beneficial agents altogether, creating the ability to release different beneficial agents at different points in time, as shown in FIG. 12. For example, in FIG. 12, a layer 60 of

anti-thrombotic agent could be deposited at the inwardly facing surface of the stent, followed by a barrier layer 52 and alternating layers of anti-proliferatives 62 and anti-inflammatories 64. This configuration could provide an initial release of anti-thrombotic agent into the bloodstream while simultaneously providing a gradual release of anti-proliferatives interspersed with programmed bursts of anti-inflammatory agents to the vessel wall. The configurations of these layers can be designed to achieve the agent delivery bursts at particular points in time coordinated with the body's various natural healing processes.

A further alternative is illustrated in FIG. 13. Here the concentration of the same beneficial agent is varied from layer to layer, creating the ability to generate release profiles of arbitrary shape. Progressively increasing the concentration of agent in the layers 66 with increasing distance from the outwardly facing surface 56, for example, produces a release profile with a progressively increasing release rate, which would be impossible to produce in a thin surface coating.

Another general method for controlling beneficial agent release kinetics is to alter the beneficial agent flux by changing the surface area of drug elution sources as a function of time. This follows from Fick's First Law, which states that the instantaneous molecular flux is proportional to surface area, among other factors:

$$J = D \frac{dC}{dx} \Rightarrow \frac{dN}{dt} = AD \frac{dc}{dx}$$

Where  $dN/dt$  is the number of molecules per unit time, A is the instantaneous drug eluting surface area, D is the diffusivity, and C is the concentration. The drug eluting surface area of a surface coated stent is simply the surface area of the stent itself. Since this area is fixed, this method of controlling release kinetics is not available to surface coated devices. Through openings, however, present several possibilities for varying surface area as a function of time.

In the embodiment of FIG. 14, beneficial agent is provided in the openings 24 in the form of microspheres, particles or the like. Individual layers 70 can then be created that contain these particles. Further, the particle size can be varied from layer to layer. For a given layer volume, smaller particle sizes increase the total



particle surface area in that layer, which has the effect of varying the total surface area of the beneficial agent from layer to layer. Since the flux of drug molecules is proportional to surface area, the total drug flux can be adjusted from layer to layer by changing the particle size, and the net effect is control of release kinetics by varying particle sizes within layers.

A second general method for varying drug eluting surface area as a function of time is to change the shape or cross-sectional area of the drug-bearing element along the axis of the opening. FIG. 15A shows an opening 70 having a conical shape cut into the material of the stent itself. The opening 70 may then be filled with beneficial agent 72 in layers as described above or in another manner. In this embodiment, a barrier layer 74 may be provided on the inwardly facing side of the opening 70 to prevent the beneficial agent 72 from passing into the blood stream. In this example, the drug eluting surface area  $A_t$  would continuously diminish (from FIG. 15A to FIG. 15B) as the bio-degradable carrier material erodes, yielding the elution pattern of FIG. 15C.

FIG. 16A shows a simple cylindrical through-opening 80 in which a preformed, inverted cone 82 of beneficial agent has been inserted. The rest of the through opening 80 is then back-filled with a biodegradable substance 84 with a much slower rate of degradation or a non-biodegradable substance, and the inwardly facing opening of the through opening is sealed with a barrier layer 86. This technique yields the opposite behavior to the previous example. The drug-eluting surface area  $A_t$  continuously increases with time between FIG. 16A and 16B, yielding the elution pattern of FIG. 16C.

The changing cross section openings 70 of FIG. 15A and the non-biodegradable backfilling techniques of FIG. 16A may be combined with any of the layered agent embodiments of FIGS. 9-14 to achieve desired release profiles. For example, the embodiment of FIG. 15A may use the varying agent concentration layers of FIG. 13 to more accurately tailor a release curve to a desired profile.

The process of preforming the beneficial agent plug 82 to a special shape, inserting in a through opening, and back-filling with a second material can yield more complex release kinetics as well. FIG. 17A shows a through opening 90 in which a spherical beneficial agent plug 92 has been inserted. The resulting

biodegradation of the sphere, in which the cross sectional surface area varies as a sinusoidal function of depth, produces a flux density which is roughly a sinusoidal function of time, FIG. 17B. Other results are of course possible with other profiles, but none of these more complex behaviors could be generated in a thin, fixed-area surface coating.

An alternative embodiment of FIGS. 18A-18C use a barrier layer 52' with an opening 96 to achieve the increasing agent release profile of FIG. 16C. As shown in FIG. 18A, the opening 24 is provided with an inner barrier layer 52 and multiple beneficial agent layers 50 as in the embodiment of FIG. 10. An additional outer barrier layer 52' is provided with a small hole 96 for delivery of the agent to the vessel wall. As shown in FIGS. 18B and 18C, the beneficial agent containing layers 50 degrade in a hemispherical pattern resulting in increasing surface area for agent delivery over time and thus, an increasing agent release profile.

FIG. 19 illustrates an alternative embodiment in which an opening in the tissue supporting device is loaded with cylindrical layers of beneficial agent. According to one method of forming the device of FIG. 19, the entire device is coated with sequential layers 100, 102, 104, 106 of beneficial agent. The interior surface 54 and exterior surface 56 of the device are then stripped to remove the beneficial agent on these surfaces leaving the cylindrical layers of beneficial agent in the openings. In this embodiment, a central opening remains after the coating layers have been deposited which allows communication between the outer surface 56 and inner surface 54 of the tissue supporting device.

In the embodiment of FIG. 19, the cylindrical layers are eroded sequentially. This can be used for pulsatile delivery of different beneficial agents, delivery of different concentrations of beneficial agents, or delivery of the same agent. As shown in FIG. 19, the ends of the cylindrical layers 100, 102, 104, 106 are exposed. This results in a low level of erosion of the underlying layers during erosion of an exposed layer. Alternatively, the ends of the cylindrical layers may be covered by a barrier layer to prevent this low level continuous erosion. Erosion rates of the cylindrical layers may be further controlled by contouring the surfaces of the layers. For example, a ribbed or star-shaped pattern may be provided on the radially inner layers to provide a uniform surface area or uniform erosion rate between the radially

inner layers and the radially outer layers. Contouring of the surfaces of layers may also be used in other embodiments to provide an additional variable for controlling the erosion rates.

While the invention has been described in detail with reference to the preferred embodiments thereof, it will be apparent to one skilled in the art that various changes and modifications can be made and equivalents employed, without departing from the present invention.

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Claims:

1. A medical device for treating cardiac arrhythmias, the device comprising:
  - a bioresorbable implantable device;
  - a plurality of openings in the implantable device; and
  - a chemically ablative agent provided in the openings, wherein the openings are configured to deliver the chemically ablative agent to tissue surrounding the expandable cylindrical device without permanently trapping any agent in the openings.
2. The medical device of Claim 1, further comprising a biodegradable barrier layer provided in the opening substantially adjacent an innermost surface of the implantable device.
3. The medical device of Claim 2, wherein the barrier layer prevents the chemically ablative agent from passing from the openings into a vessel lumen.
4. The medical device of Claim 1, wherein the chemically ablative agent is provided in the openings in a biodegradable matrix.
5. The medical device of Claim 1, wherein the openings are through openings.
6. The medical device of Claim 4, wherein the bioerodible material of the implantable device erodes at a rate slower than a rate of erosion of the biodegradable matrix.
7. The medical device of Claim 1, wherein the chemically ablative agent is a cytotoxic agent.
8. The medical device of Claim 1, wherein the chemically ablative agent is provided in the openings in a plurality of layers configured to release the

chemically ablative agent at a programmed release rate and over a desired administration period.

9. The medical device of Claim 1, further comprising a biodegradable cap layer provided in the opening substantially adjacent an outermost surface of the implantable device to prevent release of the chemically ablative agent during delivery of the device to an implantation location.

10. The medical device of Claim 1, wherein the implantable device is a stent.

11. A method of treating atrial fibrillation comprising:  
determining an ablation location within a blood vessel of a patient where ablation is desired;  
implanting a resorbable stent at the ablation location;  
creating a lesion in the blood vessel wall with the stent which electrically isolates portions of tissue; and  
allowing the stent to be completely resorbed over time.

12. The method of Claim 11, wherein the stent includes a chemically ablative agent.

13. The method of Claim 12, wherein the chemically ablative agent is located within openings in the stent.

14. The method of Claim 13, wherein a biodegradable barrier layer is provided in the openings substantially adjacent an innermost surface of the implantable device to substantially prevent delivery of the chemically ablative agent into the vessel lumen.

15. The method of Claim 13, wherein the openings are through openings.
16. The method of Claim 15, wherein the stent is formed of a bioresorbable material which erodes at a rate slower than a rate of erosion of a biodegradable matrix within the openings.
17. The method of Claim 12, wherein the chemically ablative agent is a cytotoxic agent.
18. The method of Claim 13, wherein the chemically ablative agent is provided in the openings in a plurality of layers configured to release the chemically ablative agent at a programmed release rate and over a desired administration period.
19. The method of Claim 13, wherein the stent has a biodegradable cap layer provided in the opening substantially adjacent an outermost surface of the stent to prevent release of the chemically ablative agent during delivery of the device to an ablation location.
20. The method of Claim 11, wherein the ablation location is determined by an electrical mapping procedure.

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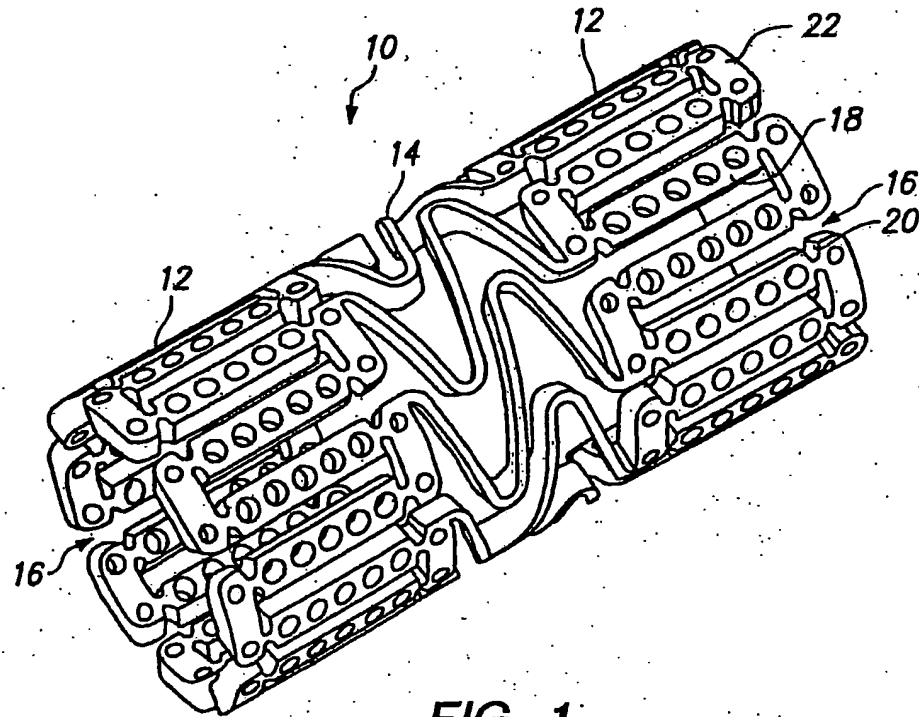


FIG. 1

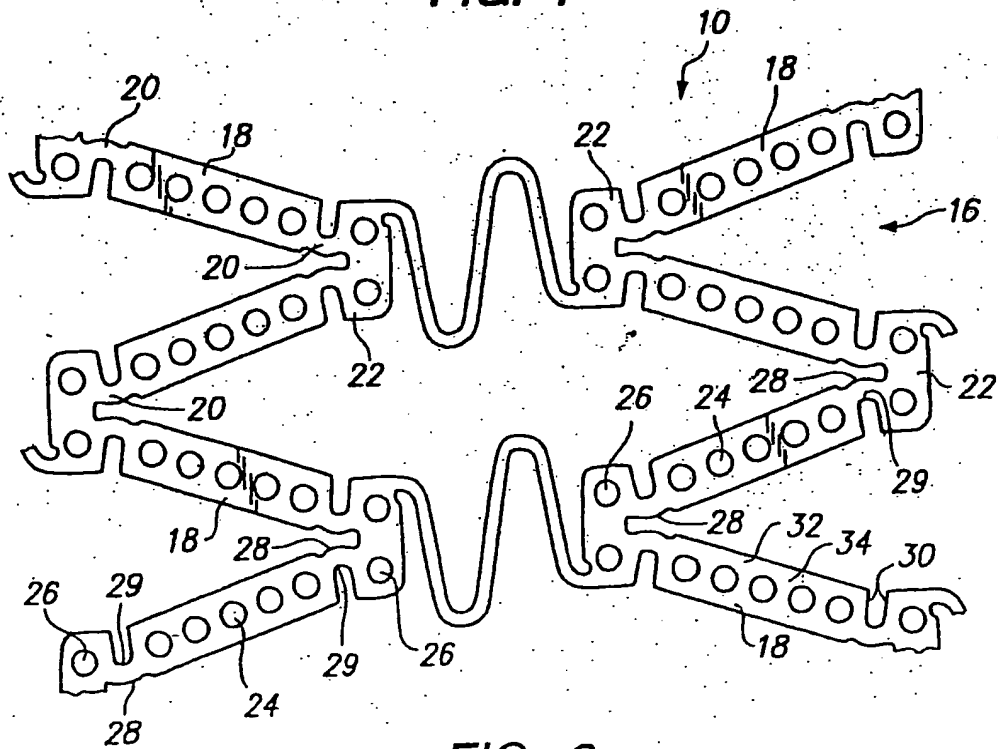


FIG. 2

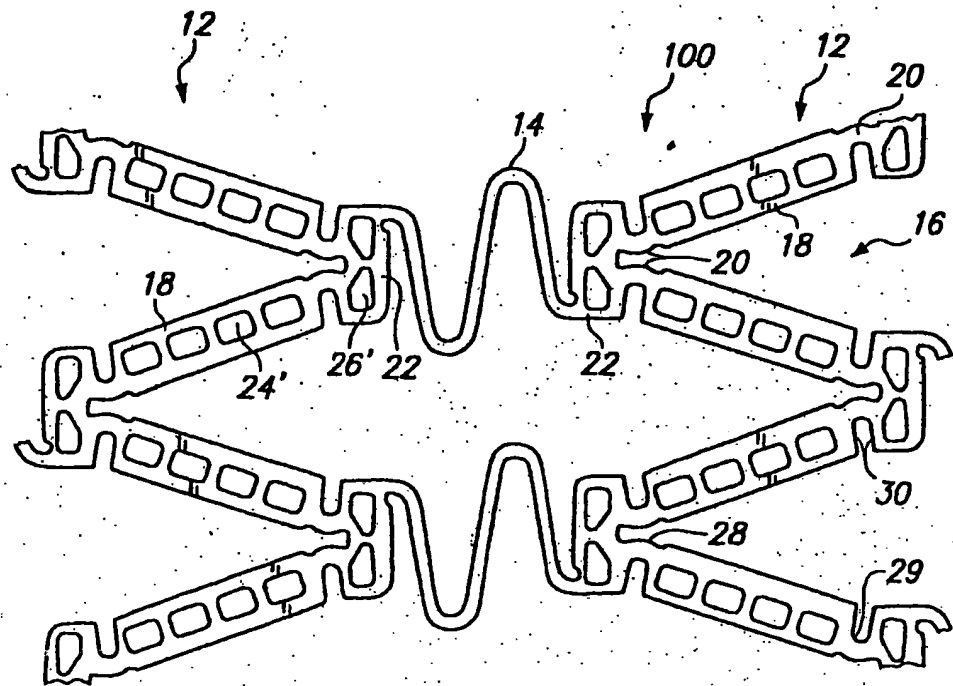
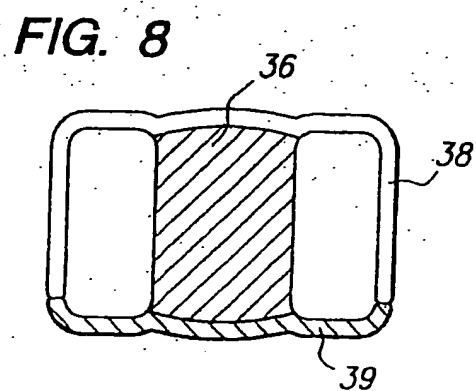
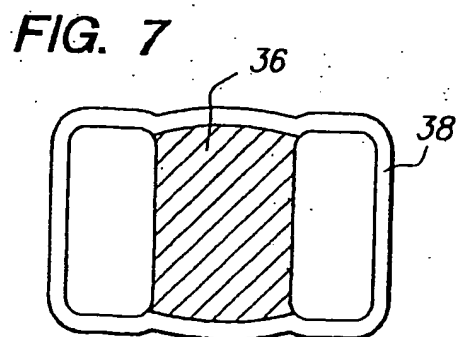
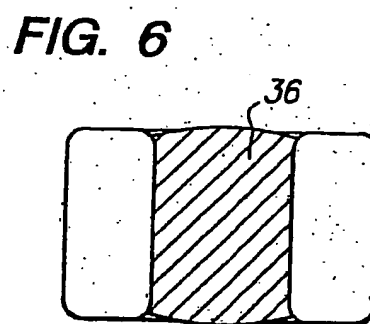
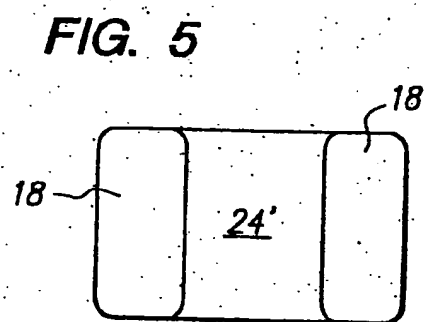
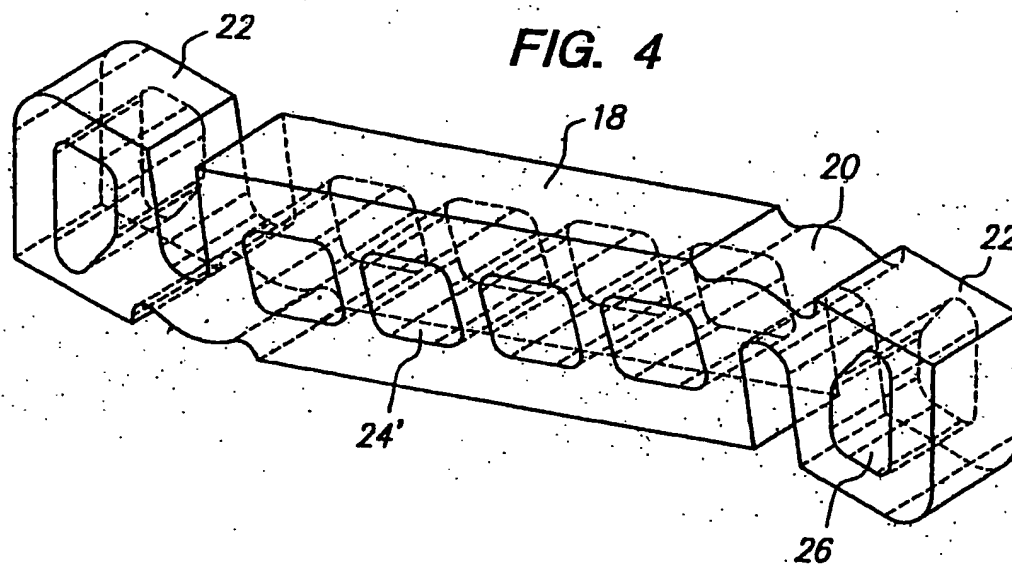


FIG. 3





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FIG. 9

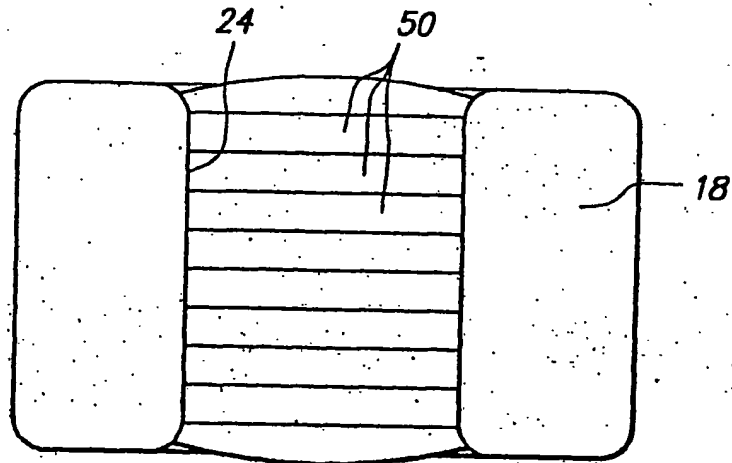


FIG. 10

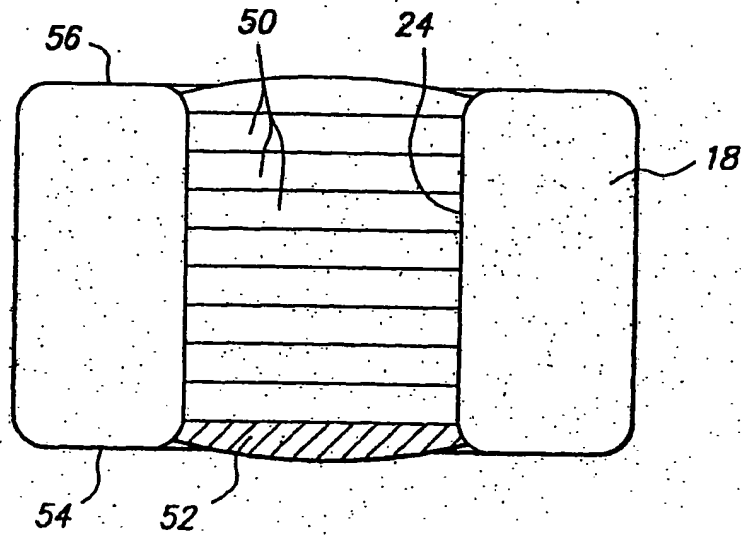
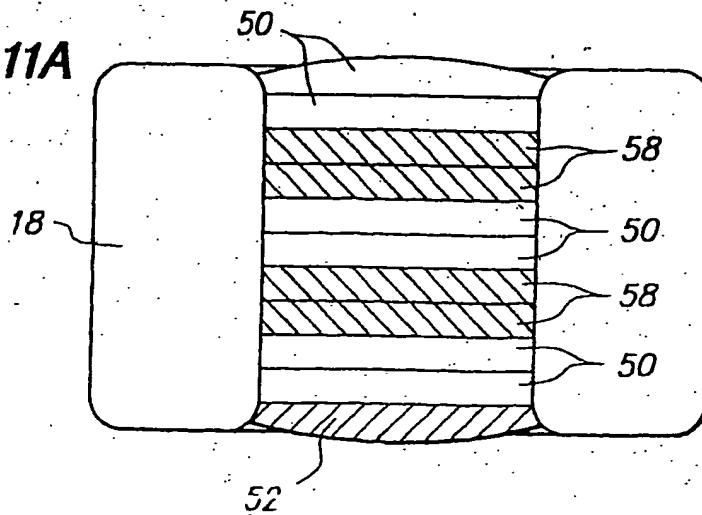
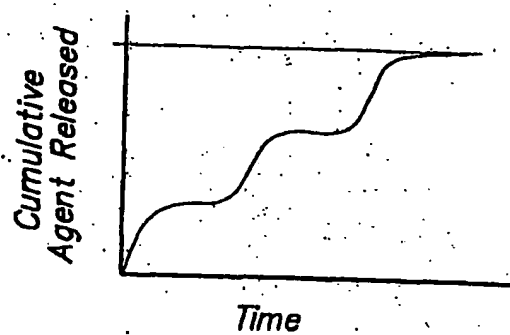


FIG. 11A

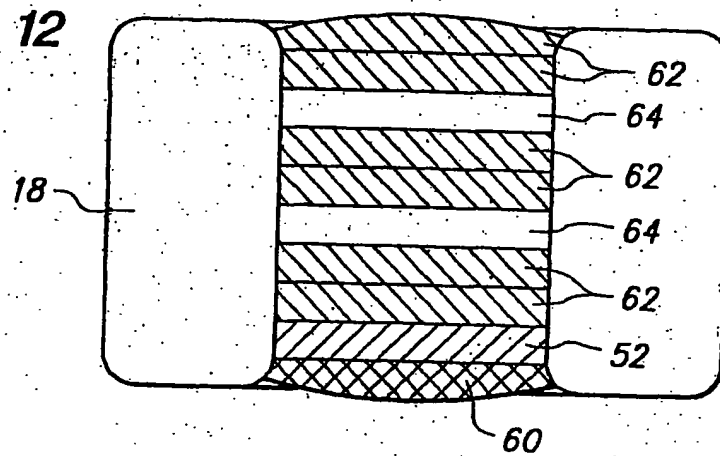


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**FIG. 11B**



**FIG. 12**



**FIG. 13**

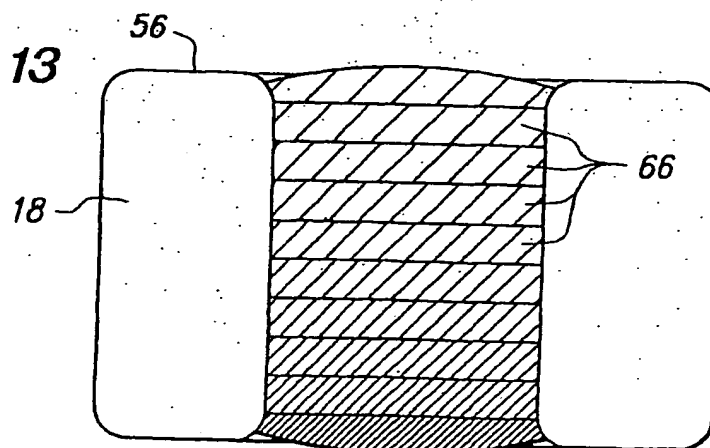


FIG. 14

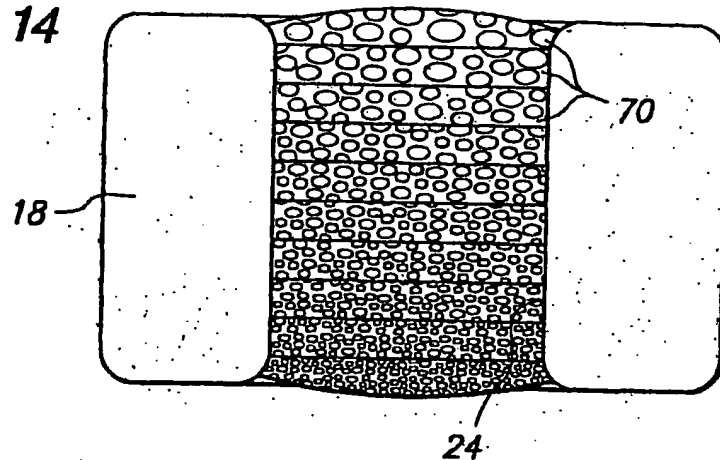


FIG. 15A

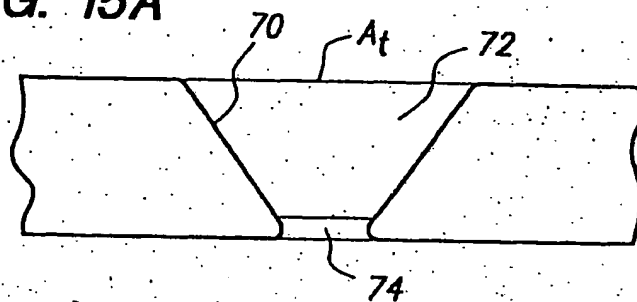


FIG. 15B

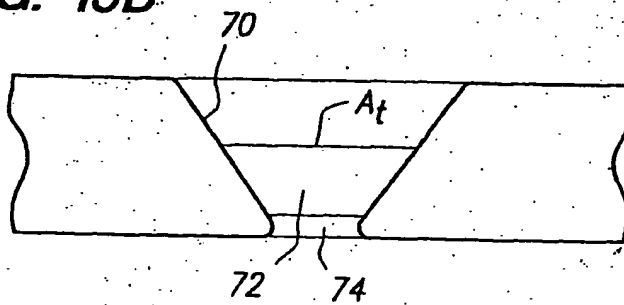
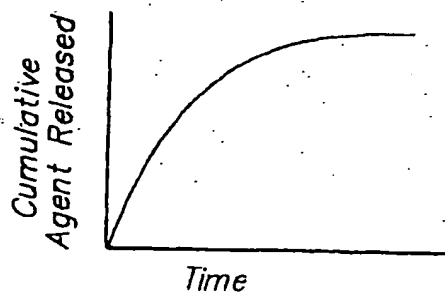
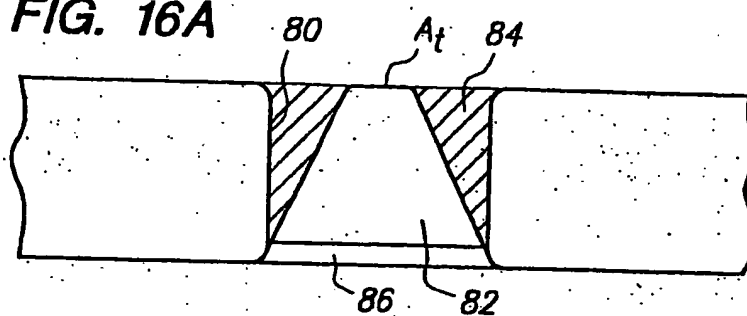
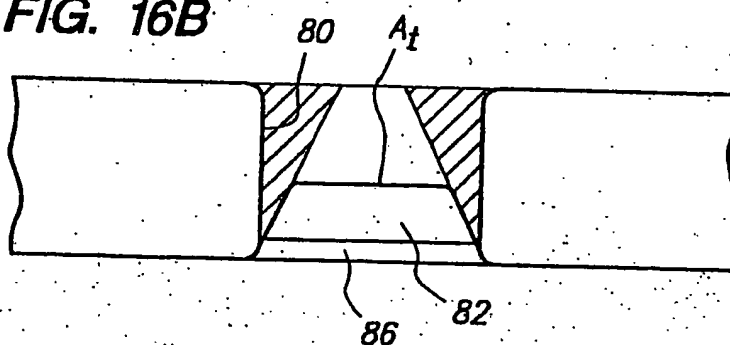
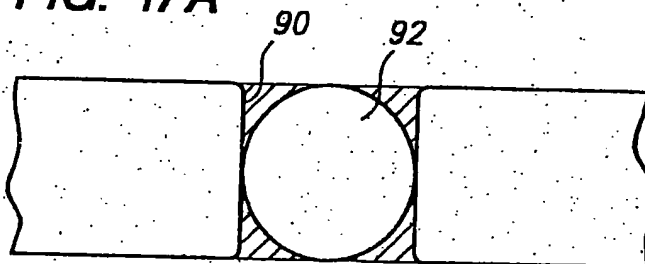
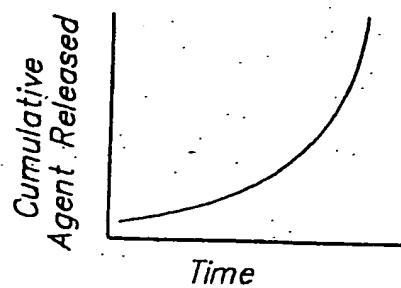
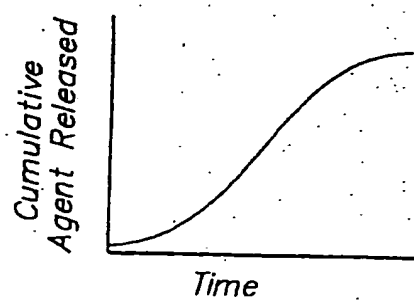


FIG. 15C



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**FIG. 16A****FIG. 16B****FIG. 17A****FIG. 16C****FIG. 17B**

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FIG. 18A

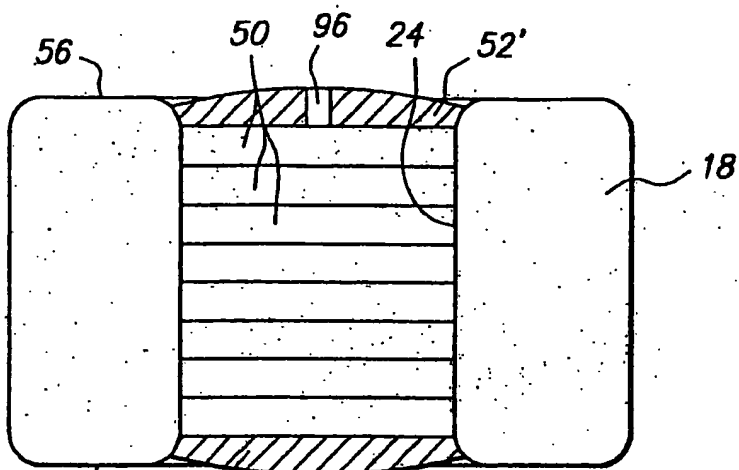


FIG. 18B

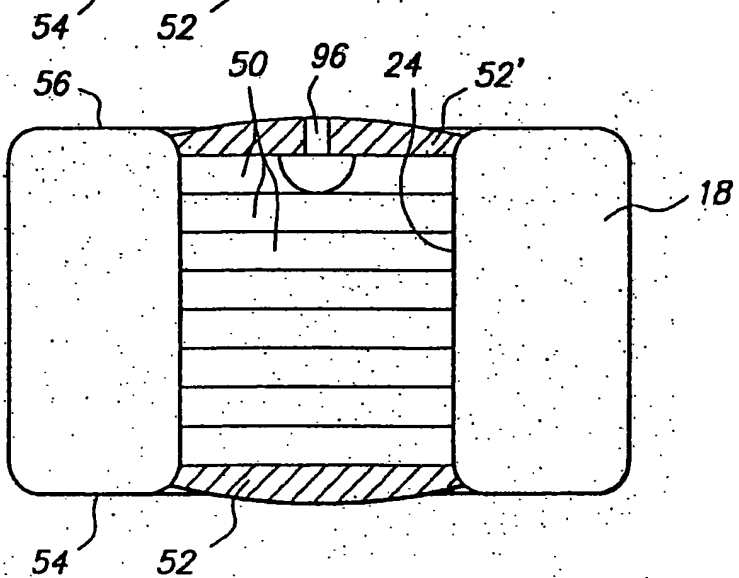
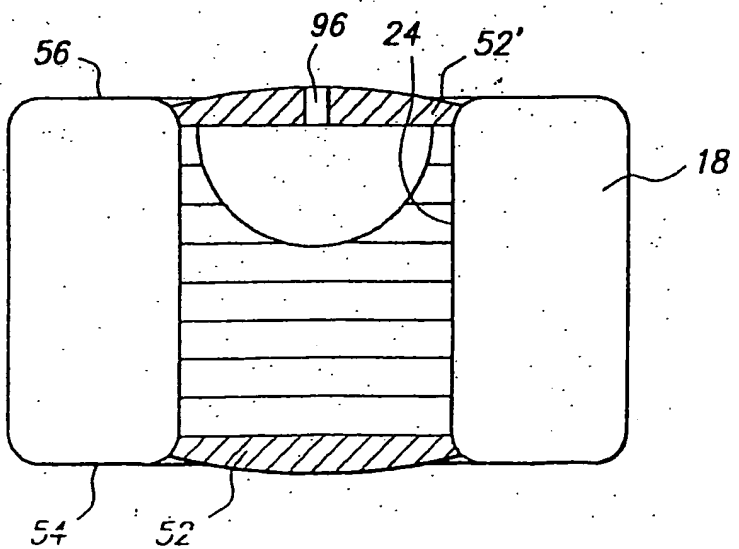
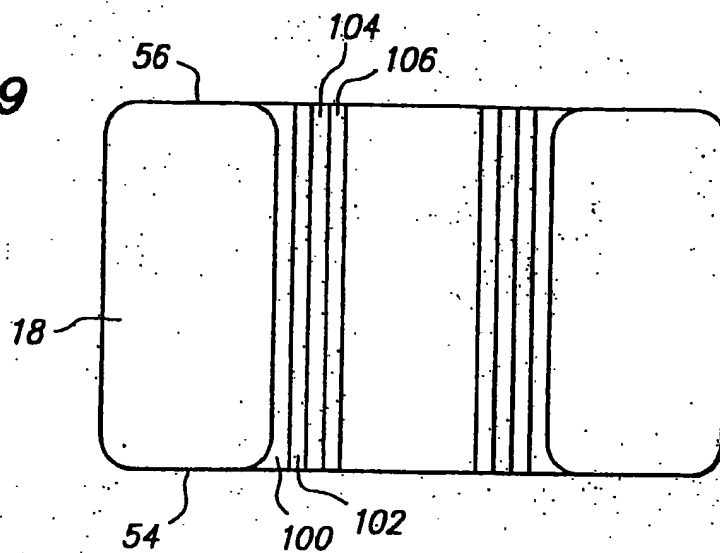


FIG. 18C



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**FIG. 19**



## INTERNATIONAL SEARCH REPORT

International application No.

PCT/US05/22243

<b>A. CLASSIFICATION OF SUBJECT MATTER</b>		
IPC(T) : A61F 2/06		
US CL 623/1.15		
According to International Patent Classification (IPC) or to both national classification and IPC		
<b>B. FIELDS SEARCHED</b>		
Minimum documentation searched (classification system followed by classification symbols) U.S. : 623/1.15, 1.21, 1.38-1.46		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)		
<b>C. DOCUMENTS CONSIDERED TO BE RELEVANT</b>		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 2003/0125803 A1 (VALLANA et al.) 03 July 2003 (03.07.2003), see entire document.	1-20
A	US 2004/0010306 A1 (FREYMAN et al.) 15 January 2004 (15.01.2004), see entire document.	1-20
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
<b>* Special categories of cited documents</b>		
"A" document defining the general state of the art which is not considered to be of particular relevance	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention	
"E" earlier application or patent published on or after the international filing date	"X" document of particular relevance, the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone	
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"T" document of particular relevance, the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art	
"O" document referring to an oral disclosure, use, exhibition or other means	"E" document member of the same patent family	
"P" document published prior to the international filing date but later than the priority date claimed		
Date of the actual completion of the international search	Date of mailing of the international search report	
03 October 2005 (03.10.2005)	27 OCT 2005	
Name and mailing address of the ISA/US Mail Stop PCT, Amd ISA/US Commissioner for Patents P.O. Box 1450 Alexandria, Virginia 22313-1450 Facsimile No. (703) 305-3230	Authorized officer Javier G. Flanco Telephone No. 571-272-4747	